

Ethnopharmacological study of plants used to treat malaria, in traditional medicine, by Bete Populations of Issia (Côte d'Ivoire)

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Abstract :

Malaria is the major tropical pathology in developing countries and man used the medicinal properties of many plants to fight this disease. During an ethnopharmacological survey in 13 villages of Issia, a Department of Côte-d'Ivoire, 37 traditional healers were interviewed. They used 32 plants to develop 32 monospecific receipts to treat malaria. Leaves and stem barks are more requested. The preparation of the medicamentous receipts utilizes mainly decoctions (81.81 %). The majority of the remedies are employed by oral way, particularly out of drink (93.93 %). For each plant, 100 g of powdered material were extracted three times in ethanol. The combined extracts were filtered and concentrated to dryness using a rotary evaporator. The ethanolic extracts were tested against chloroquine-resistant FcB1/ Colombia strain of *Plasmodium falciparum*. Extracts were prepared in DMSO at a concentration of 10 mg/ml and serially diluted with culture medium before to be added to asynchronous parasite cultures (1% antiparasitemia and 1% final hematocrite) in 96-well microplates. The growth inhibition for each extract concentration was determined by comparison of the radioactivity incorporated into the treated culture with that in the control culture maintained on the plate. The results of plants extracts evaluation showed that fourteen plants (43.75 %) had good antiplasmodial activity *in vitro* with IC₅₀ values ranging from 2.3 to 14.1 µg/ml. The ethnopharmacological informations and the results *in vitro* of antiplasmodial activities of the selected plants indicate the rightfulness of the traditional use of the studied plants as antipaludics.

Keywords: *Issia, Côte-d'Ivoire, Ethnopharmacology, Medicinal plants, Traditional Healers*

1. Introduction

Malaria is the major tropical pathology in the tropical world. A dramatic recrudescence of this disease is ongoing due to the increasing resistance of parasites. According to World Health Organization estimations [1], malaria is a several disease causing 300 to 500 millions of cases and 1.1 million deaths per year in tropical zones worldwide. Despite advances in modern medicine, malaria remains a disease which is difficult to be eradicated and is therefore a major health problem, for one main reason: all anti-malarial drugs are expensive for the populations in endemic countries [2]. In the search of means of fighting, people recognized and used the medicinal properties of many plants to fight this disease. Indeed, indigenous plants play an important role in the treatment of many diseases and 80% of people in Africa and in most of the developing countries are estimated to use herbal remedies [3]. Plants' properties are empirically appreciated despite the lack of data concerning their efficiency and safety. Also, many urban populations turn to

treatments using plants. The reason is that traditional medicine is a medicine of proximity, less constraining and non expensive [4, 5]. In the search of fighting means against malaria, ethnopharmacological investigations were conducted in Africa and in most of the developing countries [6, 7, 8, 9, 10]. Bete people in Issia, recognized the medicinal virtues of several species used to treat malaria. During an ethnopharmacological survey in Issia, Central-West of Côte d'Ivoire, 32 plants were selected, according to their commonly use by traditional healers for treatment of malaria [11]. This study aims at presenting the botanical and ethnomedical characteristics of selected medicinal plants and evaluating their *in vitro* antimalarial activities.

2. Materials and methods

2.1. Study site

Our investigations took place in villages of Issia Department (figure 1). Located at 360 km of Abidjan, Issia is part of the Central-West forest of Côte-d'Ivoire (West Africa), in the guinea field of the mesophilic sector, characterized by dense moist semi-deciduous forest [12].

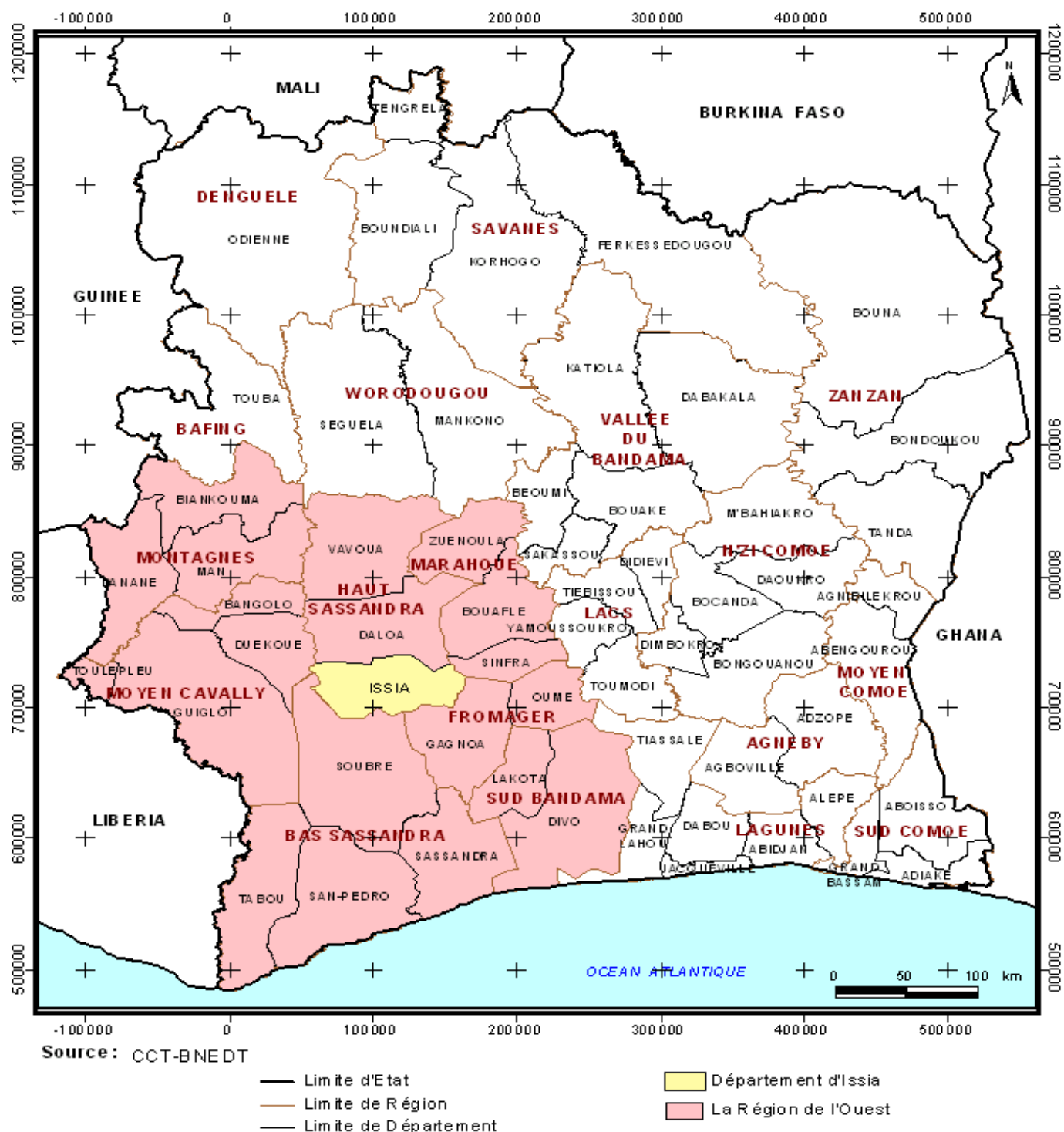


Fig. 1: Geographical situation of the Department of Issia in Côte d'Ivoire.

region is warm and humid and characterized by two seasons: a dry season from December to March and a long rainy season from April to November. The Department of Issia had 105 000 inhabitants. Bete is the native people of

Issia; there is a community of non-natives coming from all the other areas of Côte-d'Ivoire and also a community of foreigners for the most part coming from the West African Sub-region and non-

African communities (French, Lebanese and Syrian).

2.2. Plants and equipment for collection

The vegetable material is represented by all the plants that are subject of this study. As technical equipment, we used a classic material that allowed us to have access to the plants and take some samples to build up a collection of dried plants.

2.3. Ethnomedicinal survey

The investigation on the traditional use of plants having antimalarial effect was conducted among 13 native villages in the Department of Issia (Côte-d'Ivoire). As approach, we met the healers and organized semi-structured interviews. Each of them was met twice, at different moments, to answer the same questions. During this ethnomedicinal investigation, we collected informations relating to the plants used to treat malaria: different parts used as drugs, methods of collection and modes of preparation and administration of the medicamentous receipts. From the collected samples and specimens of the herbarium of the National Floristic Center (C.N.F.), we identified the plants, by their scientific name and we determined their botanical characteristics.

2.4. Preparation of plants extracts

Part of collected plants were air-dried during 7 or 10 days in the shade at room temperature and powdered. For each plant, 100g of material were extracted three times in ethanol at room temperature. The combined extracts were filtered and concentrated to dryness using a rotary evaporator. Dry extracts were stored at 4°C until analysis.

2.5. In vitro antiplasmodial activity

Extracts were tested against chloroquine-resistant FcB1/ Colombia strain of *Plasmodium falciparum* (IC₅₀ value for chloroquine of 0.1µm [13]. The antiplasmodial activity was determined according to Desjardins *et al.* (1979). Extracts were prepared in DMSO at a concentration of 10 mg/ml and serially diluted with culture medium before to be added to asynchronous parasite cultures

(1% antiparasitemia and 1% final hematocrite) in 96-well microplates. Plates were maintained for 24h at 37°C. 0.5 µCi of ³H hypoxanthine was then added to each well and parasites were maintained for further 24 h. The growth inhibition for each extract concentration was determined by comparison of the radioactivity incorporated into the treated culture with that in the control culture maintained on the plate. The concentration causing 50% inhibition of parasite growth (IC₅₀) were calculated from the drug concentration-response curves.

3. Results and discussion

3.1. Botanical characteristics of the studied plants

The ethnopharmacological investigations that we conducted in Issia (Côte-d'Ivoire), made it possible to identify 32 species of plants used in traditional medicine to treat malaria. These species of plants (table 1) belong to 19 families, 13 orders, 05 sub-classes, 1 class (Dicotyledon). The families of Apocynaceae and Euphorbiaceae, with respectively 05 and 04 individuals, are the best represented. From the viewpoint of Morphological Types, taxons can be divided into 4 groups (table 2): trees, shrubs, lianas and herbs. The trees, 13 of them, representing 40.62%, are mainly used. Concerning the Biological Types, the Phanerophytes are in the majority (92.80%). Among Phanerophytes, the Microphanerophytes are the best represented. Thirty-one species (96.87 %) of our listed plants are wild taxons; only one of them is introduced. The intensive wild plants sampling cause regression of spontaneous wild species.

3.2. Ethnopharmacological survey and characteristics of the studied plants

During this ethnopharmacological study conducted in the Department of Issia (Côte d'Ivoire), we met 37 traditional healers, native of 13 villages who accepted our collaboration and gave informations on anti-malarial plants (table 3). The ethnopharmacological characteristics of the plants, the different parts used as

drugs, the methods of preparation and administration of medicamentous receipts, are consigned in table 4. We note that 32 medicamentous receipts are developed to treat malaria by Bete people in Issia. All the receipts are monospecific. These results are similar to that of Ouattara [14] who indicated that all the medicinal formulas are monospecific. The following organs are used as drugs: leaf, stem bark, root bark and whole plant. There are different methods of sampling these organs: for roots and whole plants, the sampling is made with a hoe; leaves are usually picked by hand; concerning the stem and roots bark, the sampling is done with machetes. Easily accessible specimens (leaves) are usually picked by hand. Methods of sampling are very important; they must prevent plant species extinction [15].

There are different modes of preparation: decoction, infusion, chewing, kneading

and roasting. In this study decoction (100%) is the most widespread method of preparation (table 4). The drink (100%) is the most used method of administration. This result is similar to [14] who showed that the decoction and the drink are applied in 100 % of cases in the Department of Divo. Of the 32 receipts, 12 (37.5 %) are made with leaves, 10 (31.25%) made with stem barks, 7 (21.87%) made with whole plants and 3 (9.375%) with root barks. This result is in the line with OUATTARA's study [14]; the author showed that the leaves are mostly used in 37.50 % of the cases. We noticed that 5 receipts are made with plants belonging to Apocynaceae Family, 4 made with Euphorbiaceae species and 3 receipts are prepared with Rubiaceae species. These Families contain alkaloids with antimalarial properties [16]. According to some studies, we notice interesting similarities.

Table 1: Systematic groups of studied plants

Plants species used	Family	Order	S/C	Classes
<i>Albizia ferruginea</i>	Fabaceae	Fabales	Rosidae	Dicotyledons
<i>Alchornea cordifolia</i>	Euphorbiaceae	Euphorbiales	Rosidae	Dicotyledons
<i>Alstonia boonei</i>	Apocynaceae	Gentinales	Asteridae	Dicotyledons
<i>Alternanthera pungens</i>	Amaranthaceae	Caryophyllales	Caryophyllidae	Dicotyledons
<i>Anthocleista djalonensis</i>	Loganiaceae	Gentinales	Asteridae	Dicotyledons
<i>Anthonotha macrophylla</i>	Caesalpiniaceae	Fabales	Rosidae	Dicotyledons
<i>Bersama abyssinica</i>	Melanthaceae	Sapindales	Rosidae	Dicotyledons
<i>Cassia alata</i>	Caesalpiniaceae	Fabales	Rosidae	Dicotyledons
<i>Chrysophyllum perpulchrum</i>	Sapotaceae	Ebenales	Dilleniidae	Dicotyledons
<i>Clerodendrum splendens</i>	Verbenaceae	Lamiales	Asteridae	Dicotyledons

<i>Cyathula prostrate</i>	Acanthaceae	Scrophulariales	Asteridae	Dicotyledons
<i>Elytraria marginata</i>	Acanthaceae	Scrophulariales	Asteridae	Dicotyledons
<i>Erythrococca anomala</i>	Euphorbiaceae	Euphorbiales	Rosidae	Dicotyledons
<i>Funtumia elastica</i>	Apocynaceae	Gentianales	Asteridae	Dicotyledons
<i>Hunteria eburnean</i>	Apocynaceae	Gentianales	Asteridae	Dicotyledons
<i>Hura crepitans</i>	Euphorbiaceae	Euphorbiales	Rosidae	Dicotyledons
<i>Irvingia gabonensis</i>	Irvingiaceae	Sapindales	Rosidae	Dicotyledons
<i>Mareya micrantha</i>	Euphorbiaceae	Euphorbiales	Rosidae	Dicotyledons
<i>Melanthera scandens</i>	Asteraceae	Asterales	Asteridae	Dicotyledons
<i>Microglossa pyrifolia</i>	Asteraceae	Asterales	Asteridae	Dicotyledons
<i>Millettia zechiana</i>	Fabaceae	Fabales	Rosidae	Dicotyledons
<i>Morinda morindoides</i>	Rubiaceae	Rubiales	Asteridae	Dicotyledons
<i>Motandra guineensis</i>	Apocynaceae	Gentianales	Asteridae	Dicotyledons
<i>Nauclea latifolia</i>	Rubiaceae	Rubiales	Asteridae	Dicotyledons

Table 1 (End): Systematic groups of studied plants

<i>Olox gambecola</i>	Olacaceae	Santalales	Rosidae	Dicotyledons
<i>Parquetina nigrescens</i>	Perplocaceae	gentianales	Asteridae	Dicotyledons
<i>Phyllanthus muellerianus</i>	Euphorbiaceae	Euphorbiales	Rosidae	Dicotyledons
<i>Physalis angulata</i>	Solanaceae	Solanales	Asteridae	Dicotyledons
<i>Rauvolfia vomitoria</i>	Apocynaceae	Gentianales	Asteridae	Dicotyledons
<i>Rhigiocarya racemifera</i>	Menispermaceae	Ranunculales	Magnoliidae	Dicotyledons
<i>Scherbournia bignoniiflora</i>	Rubiaceae	rubiales	Rosidae	Dicotyledons

<i>Zanthoxylum gillettii</i>	Rutaceae	Sapindales	Rosidae	Dicotyledons
Total: 32 Species	19 Families	13 Orders	05 S/Classes	01 Class

Table 2: Morpho-Biological Types and Chorological Affinities of listed plants

Plants species used	Morphological Types	Biological Types	Chorological affinities	Status
<i>Albizia ferruginea</i>	Tree	Microphanerophyte	GC	Wild
<i>Alchornea cordifolia</i>	Shrub	Microphanerophyte	GC	Wild
<i>Alstonia boonei</i>	Tree	Megaphanesophyte	GC-SZ	Wild
<i>Alternanthera pungens</i>	Herb	Chamephyte	GC-SZ	Wild
<i>Anthocleista djalonensis</i>	Tree	Microphanerophyte	GC	Wild
<i>Anthonotha macrophylla</i>	Tree	Microphanerophyte	GC	Wild
<i>Bersama abyssinica</i>	Tree	Microphanerophyte	GC	Wild
<i>Cassia alata</i>	Shrub	Nanophnerophyte	GC	Wild
<i>Chrysophyllum perpulchrum</i>	Tree	Mesophanerophyte	GC	Wild
<i>Clerodendrum splendens</i>	Liana	Microphanerophyte	GC	Wild
<i>Cyathula prostrate</i>	Herb	Nanophanerophyte	GC-SZ	Wild
<i>Elytraria marginata</i>	Herb	Chamephyte	GC	Wild
<i>Erythrococca anomala</i>	Shrub	Nanophnerophyte	GC	Wild
<i>Funtumia elastica</i>	Tree	Mesophanerophyte	GC	Wild
<i>Hunteria eburnean</i>	Tree	Microphanerophyte	GC	Wild
<i>Hura crepitans</i>	Tree	Mesophanerophyte	I	Cultivated

<i>Irvingia gabonensis</i>	Tree	Megaphanerophyte	GC	Wild
<i>Mareya micrantha</i>	Tree	Nanophnerophyte	GC	Wild
<i>Melanthera scandens</i>	Liana	Nanophnerophyte	GC	Wild
<i>Microglossa pyrifolia</i>	Shrub	Microphanerophyte	GC	Wild
<i>Millettia zechiana</i>	Tree	Microphanerophyte	GC	Wild

Signification of Symbols: I: Introduced; GC: Guineo-Congolais; SZ: Soudano-Zambesienne

Table 2 (End): Morpho-Biological Types and Chorological Affinities of listed plants

<i>Morinda morindoides</i>	Liana	Microphanerophyte	GC	Wild
<i>Motandra guineensis</i>	Liana	Microphanerophyte	GC-SZ	Wild
<i>Nauclea latifolia</i>	Liana	Microphanerophyte	GC	Wild
<i>Olox gambecola</i>	Herb	Nanophnerophyte	GC	Wild
<i>Parquetina nigrescens</i>	Liana	Microphanerophyte	GC	Wild
<i>Phyllanthus muellerianus</i>	Shrub	Nanophanerophyte	GC	Wild
<i>Physalis angulata</i>	Herb	Therophyte	GC-SZ	Wild
<i>Rauwolfia vomitoria</i>	Tree	Microphanerophyte	GC-SZ	Wild
<i>Rhigiocarya racemifera</i>	Liana	Microphanerophyte	GC	Wild
<i>Scherbournia bignoniiflora</i>	Liana	Microphanerophyte	GC	Wild
<i>Zanthoxylum gillettii</i>	Tree	Mesophanerophyte	GC	Wild

Table 3: Overview of interviewed traditional healers per visited village

Villages	Traditional healers (Number)	Sex		Age (years)
		Male	Female	
Takouahio	03	02	01	45-56
Bobam	03	01	02	56-63
Broma	02	00	02	56-66

Dahira	03	01	02	34-46
Digbam	04	03	01	55-60
Godoua	03	02	01	30- 55
Louoboua	03	03	00	38-40
Madia	02	00	02	43-48
Nahio	03	02	01	53-58
Sereguhé	02	01	01	43-56
Tapéguia	04	02	02	18-60
Tézié	03	01	02	27-30
Zadia	02	02	00	50-60
Total: 13 villages	37	20	17	30-66

Table 4: Indications on the methods of preparation and administration of medicines

Plants species used	Part used	Mode of Preparation	Medicamentous Form	Mode of Administration
<i>Albizia ferruginea</i>	Leaf	Decoction	Decocte	Drink
<i>Alchornea cordifolia</i>	Leaf	Decoction	Decocte	Drink
<i>Alstonia boonei</i>	Stem bark	Decoction	Decocte	Drink
<i>Alternanthera pungens</i>	Whole plant	Decoction	Decocte	Drink
<i>Anthocleista djalonenensis</i>	Stem bark	Decoction	Decocte	Drink
<i>Anthonotha macrophylla</i>	Stem bark	Decoction	Decocte	Drink
<i>Bersama abyssinica</i>	Leaf	Decoction	Decocte	Drink
<i>Cassia alata</i>	Leaf	Decoction	Decocte	Drink
<i>Chrysophyllum perpulchrum</i>	Stem bark	Decoction	Decocte	Drink
<i>Clerodendrum splendens</i>	Whole plant	Decoction	Decocte	Drink
<i>Cyathula prostrate</i>	Whole plant	Decoction	Decocte	Drink

<i>Elytraria marginata</i>	Whole plant	Decoction	Decocte	Drink
<i>Erythrococca anomala</i>	Leaf	Decoction	Decocte	Drink
<i>Funtumia elastica</i>	Stem bark	Decoction	Decocte	Drink
<i>Hunteria eburnean</i>	Stem bark	Decoction	Decocte	Drink
<i>Hura crepitans</i>	Stem bark	Decoction	Decocte	Drink
<i>Irvingia gabonensis</i>	Stem bark	Decoction	Decocte	Drink
<i>Mareya micrantha</i>	Leaf	Decoction	Decocte	Drink
<i>Melanthera scandens</i>	Whole plant	Decoction	Decocte	Drink
<i>Microglossa pyrifolia</i>	Root bark	Decoction	Decocte	Drink
<i>Millettia zechiana</i>	Stem bark	Decoction	Decocte	Drink

Table 4 (End): Indications on the methods of preparation and administration of medicines

<i>Morinda morindoides</i>	Leaf	Decoction	Decocte	Drink
<i>Motandra guineensis</i>	Leaf	Decoction	Decocte	Drink
<i>Nauclea latifolia</i>	Root bark	Decoction	Decocte	Drink
<i>Olax gambecola</i>	Whole plant	Decoction	Decocte	Drink
<i>Parquetina nigrescens</i>	Leaf	Decoction	Decocte	Drink
<i>Phyllanthus muellerianus</i>	Leaf	Decoction	Decocte	Drink
<i>Physalis angulata</i>	Whole plant	Decoction	Decocte	Drink
<i>Rauwolfia vomitoria</i>	Root bark	Decoction	Decocte	Drink
<i>Rhigiocarya racemifera</i>	Leaf	Decoction	Decocte	Drink
<i>Scherbournia bignoniiflora</i>	Leaf	Decoction	Decocte	Drink
<i>Zanthoxylum gillettii</i>	Stem bark	Decoction	Decocte	Drink

Table 5: *In vitro* antiplasmodial activity of ethanol extracts of plants on *Plasmodium falciparum*

Plants species	<i>Plasmodium falciparum</i>, IC₅₀ (µg/ml)
<i>Albizia ferruginea</i>	>50
<i>Alchornea cordifolia</i>	22.3 ± 3.3
<i>Alstonia boonei</i>	12.3 ± 0.2
<i>Alternanthera pungens</i>	>50
<i>Anthocleista djalonensis</i>	>50
<i>Anthonotha macrophylla</i>	43.2 ± 0.3
<i>Bersama abyssinica</i>	20.7 ± 3.3
<i>Cassia alata</i>	>50
<i>Chrysophyllum perpulchrum</i>	12.8 ± 4.2
<i>Clerodendrum splendens</i>	>50
<i>Cyathula prostrate</i>	12.4 ± 3.8
<i>Elytraria marginata</i>	>50
<i>Eryhrococca anomala</i>	13.1 ± 3.0
<i>Funtumia elastica</i>	3.6 ± 0.9
<i>Hunteria eburnean</i>	2.2 ± 0.9
<i>Hura crepitans</i>	>50
<i>Irvingia gabonensis</i>	22.1 ± 0.8
<i>Mareya micrantha</i>	>50
<i>Melanthera scandens</i>	20.7 ± 0.4
<i>Microglossa pyrifolia</i>	31.1 ± 3.0
<i>Millettia zechiana</i>	14.1 ± 3.1
<i>Morinda morindoides</i>	9.8 ± 0.3

IC₅₀ values are the mean ± the standard deviations from three independent experiments

Table 5 (End): *In vitro* antiplasmodial activity of ethanolic extracts of plants on *Plasmodium falciparum*.

<i>Motandra guineensis</i>	16.3 ± 1.8
<i>Nauclea latifolia</i>	7.3 ± 1.3
<i>Olex gambecola</i>	5.2 ± 0.4
<i>Parquetina nigrescens</i>	20.3 ± 0.9
<i>Phyllanthus muellerianus</i>	10.3 ± 0.7
<i>Physalis angulata</i>	7.9 ± 0.3
<i>Rauvolfia vomitoria</i>	2.5 ± 0.8
<i>Rhigiocarya racemifera</i>	>50
<i>Scherbournia bignoniiflora</i>	24.7 ± 0.8
<i>Zanthoxylum gillettii</i>	2.8 ± 1.0

Some plants in this study are used by Bulu people from Cameroun in the treatment of malaria: this is the case of *Alstonia boonei* (Apocynaceae), *Morinda morindoides* (Rubiaceae) and *Rauvolfia vomitoria* (Euphorbiaceae) [17]. In Democratic Republic of Congo, people use the decoction of *Alchornea cordifolia* (Euphorbiaceae), *Morinda morindoides* (Rubiaceae) and *Rauvolfia vomitoria* (Apocynaceae) leaves in the treatment of malaria [18]; in the Mont Koupe region in Cameroun, one uses stem barks decoction of *Alstonia boonei* (Apocynaceae) and *Zanthoxylum gillettii* (Rutaceae) for their anti-malarial effects [19].

3.3. *In vitro* antiplasmodial activity of the studied plants

Of the 32 plants extracts tested, 9 (28.12%) were considered inactive against *Plasmodium falciparum* in culture with IC₅₀ values > 50 µg/ml, nine (28.12%) showed a weak antiplasmodial activity with IC₅₀ values between 15 and 50 µg/ml and fourteen (43.75%) had good *in vitro* antiplasmodial activity with IC₅₀ values ranging from 2.3 to 14.1 µg/ml (table 5). Many plants included in this study have

been described as having antiplasmodial activity; this was the case of *Funtumia elastica*, *Zanthoxylum gillettii* [20], *Morinda morindoides* [21], *Pycnanthus angolensis* [22] and *Physalis angulata* [23].

4. Conclusion

The ethnomedicinal investigations conducted in the Department of Issia (Côte-d'Ivoire) show that 32 species of plants are used by Bete people for the treatment of malaria. The drugs (stem barks, root barks and leaves) are used to develop many medicinal preparations by decoction which is the often used methods. The drinking is the most widespread mode of administration. According to some studies, we found similarities with many plants with antimalarial effects. The results of plants extracts evaluation showed that fourteen plants (43.75 %) had good *in vitro* antiplasmodial activity with IC₅₀ values ranging from 2.3 to 14.1 µg/ml. The ethnopharmacological informations and the results of *in vitro* antiplasmodial activities of the selected plants indicate the rightfulness of the traditional use of the studied plants as antipaludics.

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