

Journal of Pharmaceutical Sciences and Research

www.jpsr.pharmainfo.in

Fingerprinting of Phytosterols from Hydroalcoholic Extr act of *Ailanthus excelsa* (Roxb.) Leaves using High Performance Thin Layer Chromatography

Ravindra C. Sutar*

*Department of Pharmacology, Sanjivani College of Pharmaceutical Education and Research, At-Sahajanandnagar, Shinganapur, Kopargaon, Maharashtra, India

ravisutarbpharm@sanjivani.org.in

Abstract

Objective: Natural remedies from medicinal plants are found to be safe and effective...Many plant species have been used in folklore medicine to treat various ailments. Standardisation of plant materials is the need of the day To study flavonoid profile of the medicinal plant. *Ailanthus excelsa* (Roxb.) using High Performance Thin Layer Chromatography (HPTLC) technique. Methods: The extracts were tested to determine the presence of various phytochmeicals like alkaloids, phenolic compounds, phytosterols, carbohydrates, glycosides, saponins, terpenoids, tannins, fixed oils, fats and protein and aminoacids (Harborne and Harborne, 1998). HPTLC studies were carried out by Harborne and Wagner et al method. Different compositions of the mobile phase for HPTLC analysis were tested in order to obtain high resolution and reproducible peaks.CAMAG HPTLC system equipped with Linomat V applicator (Switzerland). Densitometric scanning was performed with Camag TLC scanner IV in the reflectance absorbance mode at 540 nm and operated by Win CATS software (1.4.6 Camag) with the help of tungstant lamp.

Results: HPTLC finger printing of phytosterols of hydroalcohol extract of leaves revealed nine polyvalent phytoconstituents (09 peaks) and corresponding ascending order of Rf values in the range of 0.010 to 0.826

Conclusion: With the results of HPTLC analysis and R_f values Phytosterols have been concluded in the extract. Hence it w as concluded that the phytosterol compounds present in the Hydroalcohol extract could be responsible for antioxidant activities. Plant derived antioxidants, especially phenols and phytosterols, have been described to have various properties like anticancer, antiaging and prevention of cardiovascular diseases. Furthur, separation and characterization of the bioactive compound from the plant is to be evaluated and reported in near future.

Keywords: HPTLC, Ailanthus excela (Roxb.) leaves, Hydroalcohol extract, Phytochemicals, Phytosterols, Fingerprinting

Introduction

Natural remedies from medicinal plants are found to be safe and effective. Many plant species have been used in folklore medicine to treat various ailments. Even today compounds from plants continue to play a major role in primary health care as therapeutic remedies in many developing countries¹. Standardisation of plant materials is the need of the day. Several pharmacopoeia containing monographs of the plant materials describe only the physicochemical parameters. Hence the modern methods describing the identification and quantification of active constituents in the plant material may be useful for proper standardisation of herbals and it's formulations. WHO has emphasized the need to ensure the quality of medicinal plant products using modern controlled technique and applying suitable standards². High performance thin layer chromatography (HPTLC) is a valuable tool for reliable identification. It can provide chromatographic fingerprints that can be visualized and stored as electronic images³. Ailanthus excels(Roxb.) (Simaroubaceae) is commonly known as Mahanimba due to its resemblance with the neem tree (Azadirachita indica) and Maharukha due to its large size. Ailanthus is from ailanto which means tree of heaven and is the name for one of the species in the Moluccas, while in Latin excelsa means tall. The plant is known by different names like tree of heaven in English⁴. Ailanthus excelsa (Roxb.) a plant used in the Indian school/system of medicine for variety of purposes⁵. Ailanthus excelsa (Roxb.) belonging to family Simaroubaceae⁶. In Chinese system of medicine bark of A. excelsa is used to treat diarrhea and dysentery, especially

when there is a blood in stool^{7,8}. Ailanthus excelsa is a fast growing tree and is extensively cultivated in many parts of India in the vicinity of villages; it is cultivated as an avenue tree for its deep shade and can be used for ant-erosion purposes⁹. The bark has been used in Asian and Australian medicine to counteract worms, excessive vaginal discharge, malaria and asthma^{10,11}. The plant shows Antifertility activity, Antifungal activity, Antimalarial activity, Hypoglycemic activity, Antipyretic activity, Antitumor and cytotoxicity, Hepatoprotective activity¹². Research interest has focused on various herbs that possess hypolipidemic, antiplatlet, antitumour, immunestimulating properties, anti-inflammatory, anti-viral etc. that may be useful adjuncts in reducing the risk of cardiovascular disease, cancer and other diseases. A wide variety of active phytochemicals, including phytosterols, terpenoids, lignans, sulfides, polyphenolics, carotenoids, coumarins, saponins, plant phytosterols, curcumins, phthalides, tannins, gallic acid, quercetin, phytophytosterols, alcohols, aldehydes have been identified from medicinal plants¹³. These phytochemicals are estimated by a variety of techniques such as spectroscopy and chromatography. High performance thin layer chromatography (HPTLC) chromatographic fingerprints can be applied for this kind of certification. Fingerprint analysis by HPTLC has developed into an effective and powerful tool for linking the chemical constituents profile of the plants with botanical identity and for estimation of chemical and biochemical markers¹⁴⁻¹⁸. Alkaloids, tannins have been Identified with HPLTC Studies of this Plant^{19,20}. but Hydroalcohol extract of this

plant has not been explored for HPTLC Studies so in this present study the HPTLC fingerprinting of Phytosterols of Hydroalcoholic extract of leaves Ailanthus excelsa (Roxb.) has been performed which may be used as markers for quality evaluation and standardization of the drug

MATERIALS AND METHODS

Plant material

Leaves of Ailanthus excelsa(Roxb.) were collected in the Month of August from the agricultural fields of Tirunelveli district, Tamilnadu. The plant was identified and leaves of Ailanthus excelsa were authenticated and confirmed from Dr.V.Chelladurai. Research Officer. Botany, C.C.R.A.S.(Retired), Govt. of India by compairing morphological features (leaf and stem arrangement, flower /inflorescence arrangement, fruit and seed morphology etc.). The collected plant material was shade dried to retain its vital phytoconstituents and then subjected to size reduction for further extraction process.

Preparation and Extraction of Plant material

Preparation of Hydroalcohol extract by Soxhlet Extraction Method: The powder of Ailanthus excelsa leaves was charged in to the thimble of a Soxhlet apparatus and extracted using Water and ethanol (1:1) proportion. Appearance of colourless solvent in the siphon tube was the indication of exhaustive extraction and based on that the further extraction was terminated. The extract was then transferred into the previously weighed empty beaker and evaporated to a thick paste on the water bath, maintained at 50°C to get Hydroalcohol extract. The extract was finally air dried thoroughly to remove all traces of the solvent and the percentage yield was calculated. The perfectly dried extract was then stored in an air tight container in a refrigerator below 10° C. The Hydroalcohol extract of Ailanthus excelsa leaves was subjected to the following investigations

1. Preliminary phytochemical screening. 2. HPTLC Fingerprinting of Phytosterols

HPTLC Fingerprinting

HPTLC studies were carried out following the method of Harborne²¹ and Wagner et al ²².

HPTLC instrumentation and Chromatographic conditions

The sample solutions were spotted in the form of bands of width 8.0 mm with a Camag microliter syringe on precoated silica gel aluminum plate 60F254 (20 cm × 10 cm with 250 um thickness; E. Merck, Darmstadt, Germany, supplied by Anchrom Technologists, Mumbai) using a Camag Linomat V (Switzerland). The plates were activated at 120°C for 20 minutes prior to chromatography. A constant application rate of 1.0 µl/s was employed, and space between two bands was 5 mm. The slit dimension was kept at $6.0 \text{ mm} \times 0.45$ mm and 10 mm/second scanning speed was employed. The slit bandwidth was set at 20 nm, each track was scanned thrice and baseline correction was used. The mobile phase for fingerprinting of phytosterols consisted of chloroformethyl acetate in the volume ratio of 4:6 (v/v), and anisaldehyde sulfuric acid was used for derivatization, 20 ml of mobile phase was used per chromatography.Linear ascending development was carried out in 20 cm × 10 cm twin trough glass chamber (Camag, Muttenz, Switzerland) saturated with filter paper whatman no: 1 in the mobile phase. The optimized chamber saturation time for mobile phase was 20 minutes at room temperature (25°C \pm 2) at relative humidity of $60\% \pm 5$. The length of the chromatogram run was 8.0 cm. Subsequent to the scanning; thin layer chromatography (TLC) plates were dried in a current of air with the help of an air dryer. Densitometric scanning was performed with Camag TLC scanner IV in the reflectance absorbance mode at 540 nm and operated by Win CATS software (1.4.6 Camag) with the help of tungsten lamp. Subsequent to the development; TLC plate was dipped in anisaldehyde sulfuric acid reagent followed by drying in the oven at 110°C. Concentrations of the compound chromatographed were determined from the intensity of diffusely reflected light. Evaluation was carried out by comparing peak areas with linear regression²³⁻³¹.

RESULTS AND DISCUSSION

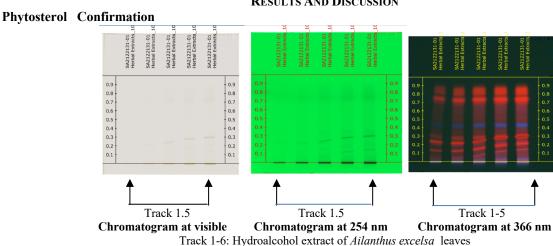


Fig. 1: HPTLC fingerprint profile of Phytosterols of leaf extract of Ailanthus excelsa Detection of Phytosterols in Hydroalcohol extract

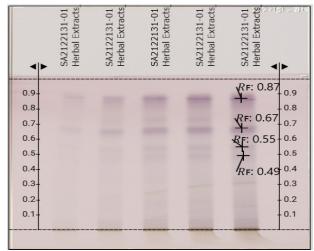


Fig. 2: Phytosterols confirmation at visible derivatisation with Anisaldehyde Sulphuric acid reagent

Table 1: Rf Values for phytosterols in Hydroalcohol extract of Ailanthus excelsa leaves

1 V											
Peak	Start		Max			End		Area		Manual	Substance
#	R _F	Н	RF	Н	%	RF	Н	Α	%	peak	Name
1	0.000	0.0000	0.010	0.2086	17.81	0.021	0.0000	0.00234	5.79	No	
2	0.060	0.0000	0.079	0.0110	0.94	0.116	0.0000	0.00041	1.01	No	
3	0.121	0.0000	0.160	0.0875	7.47	0.181	0.0116	0.00159	3.95	No	
4	0.182	0.0115	0.226	0.1137	9.71	0.273	0.0073	0.00455	11.27	No	
5	0.273	0.0073	0.315	0.3992	34.09	0.356	0.0072	0.00945	23.40	No	
6	0.410	0.0000	0.435	0.0106	0.90	0.484	0.0000	0.00041	1.02	No	
7	0.485	0.0000	0.600	0.0415	3.54	0.652	0.0184	0.00377	9.34	No	
8	0.660	0.0179	0.750	0.2011	17.17	0.792	0.0642	0.01286	31.85	No	
9	0.794	0.0640	0.826	0.0980	8.37	0.852	0.0854	0.00499	12.36	No	

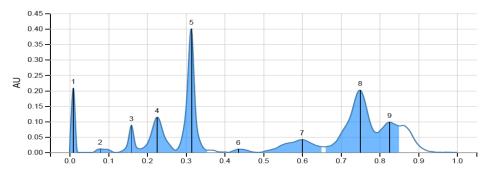


Fig 3: Chromatogram for phytosterols in Hydroalcohol extract of Ailanthus excelsa leaves

It was observed that track 1-5 (Figure 1) shows Hyderoalcohol extract. Figure 2, the fluorescence shows purple bands which concludes the presence of Phytosterols in the extract. It was observed that there is a separation of different phytoconstituents, in Hydroalcohol extract

Fingerprinting study of Hydroalcohol extract at 366 nm shows nine (09 Peaks) Rf between the range of 0.010-0.826. Rf 0.315 has maximum 34.09 % concentration in Table 1, Figure 3 shows R_f Values for Phytosterols in Hydroalcohol extract of *Ailanthus excelsa* leaves.

The evaluation of crude extract is an integral part of correct identity. HPTLC is useful as a phytochemical marker^[32, 33] and more effective in the field of plant taxonomy also for the identification of plants through secondary metabolites³⁴. HPTLC fingerprinting is proved to be a linear, precise, and accurate method for herbal identification³⁵. Such finger printing is useful in quality control of herbal products and

checking for the adulterants³⁶. Therefore, it can be useful for the evaluation of different marketed pharmaceutical preparations ^{37–39}.

Due to the adverse effects of synthetic drugs, in recent vears, scientists are on the search for alternative medicine. There are some diseases which are chronic and need a long duration of medication, plant-based drugs are less toxic and have no side effects. We have got positive results for antiarthritic activity of this plant in our previous studies. Furthermore, the literature survey of phytosterols has shown potent antiarthritic, antiinflammatory, activity40-44. immunosuppressant In recent years, phytosterols like beta sitosterol have shown central inhibitory and neuromodulatory functions which claims its as an anxiolytic, sedative, anticonvulsant, antidepressant activities in our studies⁴⁵.

CONCLUSION

It is observed in the above HPTLC studies that, Hydroalcohol extract of *Ailanthus excelsa* (Roxb.) contain a lot of polyvalent chemical constituents with different R_f values. The developed fingerprint analysis of leaf extract of *Ailanthus excelsa* will help to isolate and identify new Phytosterols which will offer a possibility to discover a lead molecule for drug development.

ACKNOWLEDGMENT

The author wish to thank Anchrom Test Lab Pvt. Ltd. Mulund (E), Mumbai - 400081 for their excellent and generous help for the HPTLC analysis.

AUTHOR CONTRIBUTIONS

Dr. Ravindra C. Sutar conceptualized and designed the study, curated the data and prepared the original draft, discussed the methodology and analysed the data, prepared results and contributed to the final manuscript.

FUNDING

I acknowledge the resource support for the study was provided by the Hon'ble Management of Sanjivani Group of Institutes, Sanjivani College of Pharmaceutical Education and Research, Kopargaon. The excellent support for carrying out the HPTLC study at Anchrom Test Lab Pvt. Ltd. Mulund (E), Mumbai – 400081, is also acknowledged.

REFERENCES:

- Bobbarala V, Bramhachari PV, Ravichand J, Reddy YHK, Kotresha D, Chaitanya KV. Evaluation of hydroxyl radical scavenging activity and HPTLC fingerprint profiling of *Aegle marmelos* (L.) Correa extracts. J Pharm Res 2011; 4(1):252-255.
- Sharma P, Kaushik S, Jain A, Sikarwar S M. Preliminary phytochemical screening and HPTLC fingerprinting of Nicotiana tobacum leaf. J Pharm Res 2010; 3(5):1144-1145.
- Johnson M, Mariswamy Y, Ganaraj WE. Chromatographic finger print analysis of steroids in *Aerva lanata* L by HPTLC technique. Asian pac j Trop Biomed 2011; 1:428-433.
- Database, 2000. Medicinal Plants Used in Ayurveda. Central Council for Research in Ayurveda and Siddha. New Dhli, India, pp: 50-59.
- Kirtikar, K.R. and B.D. Basu, 1995. Indian Medicinal Plants. Vol. 1, International Book Distributors, Dehradun, India, 1995; 371-372.
- Anonymous. The Wealth of India, Raw Materials. Publication and information Directorate, New Delhi, 1985:116-118.
- Chopra, R.N., I.C. Chopra, K.L. Handa and L.D. Kapur, 1958. Chopra's Indigenous Drugs of India. 2nd Edn.,UN. Dhar and Sons Private Ltd., Calcutta, 1958: 408.
- Dash, S.K. and S. Padhy. Review on ethnomedicines for diarrhoea diseases from *Orissa*: Prevalence versus culture. J. Hum. Ecol., 2006.20: 59-64.
- Anonymous, 1956. The Wealth of India: Raw Materials. Council of Industrial and Scientific Research, New Delhi.
- Kirtikar, K.R. and B.D. Basu, 2003. Indian Medicinal Plant. 2nd Edn., Mohan Basu Publisher, Allahabad, India.
- Chevallier, A., 1996. The Encyclopedia of Medicinal Plants. 1st Edn., DK Publishing Inc., New York, USA.:259.
- Lavhale, M.S. and S.H. Mishra, 2007. Nutritional and therapeutic potential of *Ailanthus excelsa*: A review. Pharmacognosy Rev., 1: 105-113.
- Craig. W, Beck.L Phytochemicals:Health protective effects. Can J Diet Pract Res. 1999; 60(2): 78-84
- Patil AG, Koli SP, Patil DA, Chandra N, Pharmacogonostical standardization and HPTLC fingerprint of Crataeva tapia Linn.SSP. Odora(Jacob.) Almeida leaves. Int.j.pharm.Biosci.2010; 1(2): 1-14

- Ramya V, Dheena Dhayalan V, Umamaheswari S. In vitro studies on antibacterial activity and separation of active compounds of selected flower extracts by HPTLC. J.Chem.Pharm.Res., 2010; 2(6): 86-91.
- Manikandan A,Victor Arokia Doss A.Evaluation of biochemical bontents, nutritional value,trace elements,SDS-PAGE and HPTLC profiling in the leaves of Ruellia tuberose L.and Dipteracanthus patulus (Jacq.).J.Chem.Pharm.Res.2010; 2(3): 295-303
- Yamunadevi M, Wesely EG, Johnson M. Chromotographic fingerprint analysis of steroids in Aerva lanata L.by HPTLC technique. Asian Pac. J. Trop. Biomed. 2011; 1: 428-433.
- Yamunadevi M, Wesely EG, Johnson M.Chemical profile studies on the alkaloids of medicinally important plant Aerva lanata L using HPTLC.J.Nat.Conscientia.2011; 2(2): 341-349.
- Ranjana Sharma, Sudhir Singh Gangwar, Amita Tilak and Ravindra C. Sutar. High performance thin layer chromatography fingerprinting of the alkaloids from *Ailanthus excelsa* (Roxb.) Leaves. WJPR.2020; 9(1):1596-1601.
- Sudhirsingh Gangwar, AmitaTilak, RanjanaSharma, Ravindra C.Sutar. HPTLC finger printing analysis of the tannins from *Ailanthus excelsa* (Roxb.) Leaves. WJPR.2018;8(2):1023-1029.
- 21. Harborne J B. *Phytochemical methods*; 3rd edition, London: Chapman and Hall: 1998.
- Wagner H, Baldt S. *Plant drug analysis*; Berlin: Springer, 1996.
 R.P.W. Scott, Encyclopedia of Chromatography, 10th edn, Marcel Dekker, USA, 2001; 252–254.
- ICH/CPMP Guidelines Q2B, Validation of Analytical Procedures— Methodology, 1996.
- J. Cazes and R.P.W. Scott, Chromatography Theory, Marcel Decker, NY, 2002; 443-454.
- 25. Reviewer Guidance, Validation of Chromatographic Methods,1994.
- P.D. Sethi, HPTLC: Quantitative Analysis of Pharmaceutical Formulations, CBS Publications, New Delhi, 1996;162–165.
- E. Heftman, Chromatography Fundamentals and Applications of Chromatography and Related Differential Migration Methods. Vol. 69A, 6th edn, Elsevier, Amsterdam. 2004;253–291.
- British Pharmacopoeia, International edn, Vol. II, HMSO, Cambridge, 2002; Appendix 112 (IB).
- J. Sherma, Encyclopedia of Pharmaceutical Technology, 2nd edn, Marcel Dekker, USA, 2001; 252–254.
- ICH/CPMP guidelines Q2A, Text on Validation of Analytical Procedures, 1994.
- USP 23, NF 19, Asian edn, United States Pharmacopeial Convention, Rockville, M.D., 982, 1225.
- M. Attimarad, K. Mueen Ahmed, B. E. Aldhubaib, and S. Harsha, "High-performance thin layer chromatography: a powerful analytical technique in pharmaceutical drug discovery," *Pharmaceutical Methods*, vol. 2, no. 2, pp. 71–75, 2011.
- 33. H. Misra, D. Mehta, B. K. Mehta, and D. C. Jain, "Extraction of artemisinin, an active antimalarial phytopharmaceutical from dried leaves of *Artemisia annua* L., using microwaves and a validated HPTLC-visible method for its quantitative determination," *Chromatography Research International*, vol. 2014, Article ID 361405, 11 pages, 2014.
- K. Salim, K. S. Rajeev, and Z. A. Malik, "Assessment of phytochemical diversity in *Phyllanthus amarus* using HPTLC fingerprints," *Indo-Global Journal of Pharmaceutical Sciences*, vol. 1, pp. 1–12, 2011.
- N. Cortés, C. Mora, K. Muñoz et al., "Microscopical descriptions and chemical analysis by HPTLC of *Taraxacum officinale* in comparison to *Hypochaeris radicata*: a solution for mis-identification," *Brazilian Journal of Pharmacognosy*, vol. 24, no. 4, pp. 381–388, 2014.
- P. Teo, F. Ma, and D. Liu, "Evaluation of Taurine by HPTLC reveals the mask of adulterated edible Bird's nest," *Journal of Chemistry*, vol. 2013, Article ID 325372, 5 pages, 2013.
- S. P. Gandhi, M. G. Dewani, T. C. Borole, and M. C. Damle, "Development and validation of stability indicating HPTLC method for determination of diacerein and aceclofenac as bulk drug and in tablet dosage form," *E-Journal of Chemistry*, vol. 9, no. 4, pp. 2023– 2028, 2012.
- S. Meena and S. M. Sandhya, "Validated HPTLC method for simultaneous analysis of pyrimethamine and sulphadoxine in pharmaceutical dosage forms," *Journal of Chemistry*, vol. 2013, Article ID 698490, 6 pages, 2013.

- K. G. Patel, N. R. Jain, and P. A. Shah, "Stability indicating HPTLC method for analysis of rifaximin in pharmaceutical formulations and an application to acidic degradation kinetic study," ISRN Analytical Chemistry, vol. 2013, Article ID 613218, 9 pages, 2013.
- Crofford LJ, Wilder RL. Arthritis and autoimmunity in animals. In: McCarty DJ, Koopman WJ, editors. Arthritis and Allied Conditions. London: Lea and Febiger; 1993. p. 525-39.
- Walz DT, DiMartino MJ, Misher A. Adjuvant-induced arthritis in rats.
 II. Drug effects on physiologic, biochemical and immunologic parameters. J Pharmacol Exp Ther 1971;178(1):223-31.
- Bouic PJ. Sterols/sterolins: The natural, nontoxic immuno-modulators and their role in the control of rheumatoid arthritis. Arthritis Trust 1998;???:3-6.
- Gupta MB, Nath R, Srivastava N, Shanker K, Kishor K, Bhargava KP. Anti-inflammatory and antipyretic activities of beta-sitosterol. Planta Med 1980;39(2):157-63.
- 44. Bouic PJ, Etsebeth S, Liebenberg RW, Albrecht CF, Pegel K, Van Jaarsveld PP. Beta-sitosterol and beta-sitosterol glucoside stimulate human peripheral blood lymphocyte proliferation: Implications for their use as an immunomodulatory vitamin combination. Int J Immunopharmacol 1996;18(12):693-700.
- 45. Aguirre-Hernández E, Rosas-Acevedo H, Soto-Hernández M, Martínez AL, Moreno J, González-Trujano ME. Bioactivity-guided isolation of beta-sitosterol and some fatty acids as active compounds in the anxiolytic and sedative effects of *Tilia americana* var. mexicana. Planta Med 2007;73(11):1148-55.