









were fitted to a simple linear equation and the fitted parameters such as slope and positive correlation coefficient were found to be 0.996 and 0.999, respectively, for both of the proteins. These observations unambiguously demonstrate that the reliability of the CIntX on calculating the  $k_{rc}$  of NHs in proteins. The program, then, calculates residue-specific free energies and protection factors of proteins with the only requirement of having NMR-derived  $k_{ex}$  of NHs in the proteins. In the third stage, the program recalculates the  $k_{rc}$  of NHs by accounting the effect of ionisable groups in proteins

and calculates  $k_{rc}$  of labile protons that are not NHs in the fourth stage of the program (discussed in the next headings).

### 3.4 Revaluation of $k_{rc}$ values of backbone labile protons in proteins after accounting the effect of ionisable side chains

In general, polypeptide chains consist of about 25% amino acids having ionisable groups (COOH/NH<sub>3</sub>/SH/OH) in their side chains [16, 17]. These ionisable groups will be fully or partially protonated depending on their pK<sub>R</sub> under defined experimental conditions. Bai *et al.* have determined the effect of protonated and deprotonated forms of a few of

**Table 3:** The percentage of protonated and deprotonated forms of ionisable groups in proteins at pD 3.6 and their dissociation constants (pK<sub>R</sub>).

Residues	pK <sub>R</sub>	% Protonated forms	% Deprotonated forms
Asparatate	3.67	54	46
Glutamate	4.25	18	82
Histidine	6.54	100	0
Cysteine	8.55	100	0
Tyrosine	9.6	100	0
Lysine	10.4	100	0
N-terminal	8.00*	100	0
C-terminal	3.67**	54	46

\*pK<sub>R</sub> of N-terminal amino group.

\*\*pK<sub>R</sub> of C-terminal carboxylic acid group.

**Table 4:** Values of  $k_{rc}$  and  $\Delta G_{HX}$  of NHs of a few amino acids in cobrotoxin at pD 3.6, 298 K have been tabulated, herein.

Sl. No.	Residue <sup>#</sup>	Uncorrected <sup>*</sup>		Corrected <sup>**</sup>		$\Delta\Delta G_{HX}$
		$k_{rc}$	$\Delta G_{HX}$	$k_{rc}$ (min <sup>-1</sup> )	$\Delta G_{HX}$	
1	E2	0.13	2.26	0.10	2.08	0.18
2	C3	1.04	3.62	0.84	3.48	0.14
3	H4	2.18	NA <sup>@</sup>	2.18	NA	NA
4	N5	2.52	NA	2.52	NA	NA
5	E21	0.31	1.4	0.23	1.22	0.18
6	T22	0.25	NA	0.20	NA	NA
7	D31	0.97	NA	0.35	NA	NA
8	H32	3.01	NA	1.32	NA	NA
9	R33	0.97	NA	0.97	NA	NA
10	E38	0.33	3.57	0.24	3.38	0.19
11	R39	0.36	2.93	0.29	2.8	0.13
12	E51	0.12	2.19	0.09	2.01	0.18
13	I52	0.06	2.23	0.05	2.11	0.12
14	D58	0.93	NA	0.33	NA	NA
15	R59	0.58	3.46	0.26	2.98	0.48
16	C-terminal <sup>§</sup>	0.26	NA	0.07	NA	NA

<sup>#</sup>Amino acids are denoted by single letter codes.

<sup>\*</sup>The  $k_{rc}$  values of NHs are calculated without accounting the effect of ionisable groups in the protein.

<sup>\*\*</sup>The  $k_{rc}$  values of NHs are calculated by accounting the effect of ionisable groups in the protein.

<sup>@</sup>NA represents the residues for which the values of  $k_{ex}$  are not available.

<sup>§</sup>C-terminal carboxylic acid group.

**Table 5:** Estimation of the  $k_{rc}$  values of labile protons in the side-chains of cobrotoxin at pD 3.6, 298K.

Sl. No.	Residue <sup>#</sup>	Side Chain Proton	Rate Constant $k_{rc}$ (sec <sup>-1</sup> )	Time Constant $\tau$ (sec)
1	N5	H <sub>E</sub>	1.47	0.68
2	Q6	H <sub>E</sub>	0.98	1.02
3	Q6	H <sub>Z</sub>	0.41	2.42
4	Q7	H <sub>E</sub>	0.71	1.40
5	Q7	H <sub>Z</sub>	0.30	3.32
6	Q10	H <sub>E</sub>	0.57	1.76
7	Q10	H <sub>Z</sub>	0.25	4.02
8	N23	H <sub>E</sub>	1.07	0.93
9	R28	N $\delta$ H	0.17	5.97
10	W29	N <sub>i</sub> H	0.56	1.79
11	R30	N $\delta$ H	0.10	10.13
12	R33	N $\delta$ H	0.86	1.17
13	R36	N $\delta$ H	0.14	7.01
14	R39	N $\delta$ H	0.25	4.01
15	N48	H <sub>E</sub>	1.53	0.66
16	N53	H <sub>E</sub>	0.76	1.32
17	R59	N $\delta$ H	0.22	4.50
18	N61	H <sub>E</sub>	0.99	1.01
19	N62	H <sub>E</sub>	2.22	0.45

<sup>#</sup> Amino acids are denoted by single letter codes.

these amino acids (Asp, Glu, and His) on the intrinsic exchange rates of labile protons [7]. It is usually assumed (in the calculations of exchange rates of NHs in proteins) that an ionisable groups will be fully protonated when its  $pK_R$  is greater than the solution pD and fully deprotonated when its  $pK_R$  is lower than the pD of the solution. However, when there is not much difference ( $< 1.0$ ) between the  $pK_R$  of an ionisable group and the solution pD, the ionisable group in a polypeptide chain will have significant amount of its protonated and deprotonated forms under the defined experimental conditions. Table 3 shows the  $pK_R$  of ionisable groups of amino acids (Asp, Glu, His, Cys, Tyr, and Lys) and their percentage of protonated/deprotonated forms at pD 3.6, 298 K. The CIntX determines the fraction of protonated ( $F_{HA}$ ) and deprotonated ( $F_A$ ) forms of those ionisable groups using equation (3) and equation (4) (refer method section) and then recalculates the  $k_{rc}$  of NHs that are influenced by the ionisable groups using the equation (15) described below herein in the third stage of the program.

$$k_{rc} = 10^{(X - pD)} + 10^{(Y + pD - pKD)} + 10^{(Z)} \quad (15)$$

where,

$$X = \log k_a + \log (F_{HA}^*A_{L+} + F_A^*A_{L-}) + \log (F_{HA}^*A_{R+} + F_A^*A_{R-}) + (E_a/4.57) * (1/293 - 1/T) \quad (16)$$

$$Y = \log k_b + \log (F_{HA}^*B_{L+} + F_A^*B_{L-}) + \log (F_{HA}^*B_{R+} + F_A^*B_{R-}) + (E_b/4.57) * (1/293 - 1/T) \quad (17)$$

$$Z = \log k_w + \log (F_{HA}^*B_{L+} + F_A^*B_{L-}) + \log (F_{HA}^*B_{R+} + F_A^*B_{R-}) + (E_w/4.57) * (1/293 - 1/T) \quad (18)$$

wherein '+' and '-' signs in the subscripts of the standard reference parameters of a residue indicate the protonated and deprotonated forms of the corresponding amino acid in the polypeptide chain, respectively [7,15]. Table 4 shows the recalculated  $k_{rc}$  values for the NHs of cobrotoxin, which are influenced by ionisable groups present in the protein. Residue-specific free energy ( $\Delta G_{HX}$ ) values of the NHs have also been calculated using NMR-derived  $k_{ex}$  values of the NHs at identical exchange conditions (pD 3.6, temperature 298K) of the protein [18]. The highest  $\Delta\Delta G_{HX}$  was found to be 0.48 kcal/mol for R59 of the protein at pD 3.6, 298 K, as the residue is preceded by D58, an amino acid having ionisable group. This observation reveals that discrepancy between the  $k_{rc}$ -corrected and the  $k_{rc}$ -uncorrected will be remarkable for NHs depending on their positions in the polypeptide chain and the exchange-conditions such as pH and temperature. In the fourth stage, the CIntX calculates  $k_{rc}$  values of labile protons present in the side chains of arginine, asparagine, glutamine and tryptophan using equation (11) in a single step. Table 5 shows  $k_{rc}$  values of labile protons from a few side chains of cobrotoxin at pD 3.6 and 298 K. The labile protons of these side-chains in the protein are exchanging in the time span of 0.5 – 10.1 seconds (Table 5). The  $k_{rc}$  values of side-chain labile protons will be powerful probes to determine the folding rate(s) of tertiary structural formations of proteins in which the labile protons are protected from the solvent exchange under native conditions.

Error-free estimation of  $k_{rc}$  values for labile protons of proteins is indispensable to estimate accurate  $\Delta G_{HX}$  ( $\Delta G_{HX} = -RT \ln (k_{ex}/k_{rc})$ ) of proteins. The exchange of free energy is important to understand the relationship between three-dimensional (3D) conformations and stabilities of proteins as each protein adopts a specific, well-defined 3D structure, which is important for its biological activities [19]. Moreover,  $\Delta G_{HX}$  analysis provides clues on understanding the mechanism of unfolding of proteins (two-state/multi-state processes) and on analysing the 3D structural architectures (domains organization) of proteins [20, 21]. Thus, it is obvious that estimation of an accurate  $\Delta G$  (free energy change) for proteins at ambient conditions is indispensable to unambiguously address the thermodynamic and kinetic events of proteins. In these backgrounds, we strongly believe that CIntX, fully automated and user-friendly computational tool facilitated with web server (<http://sblab.sastru.edu/cintx.html>), is very useful for estimating  $k_{rc}$  values of all types of labile protons (backbone, side chains, N-terminal and C-terminal positions) of proteins.

#### 4. CONCLUSION

We have herein developed a software tool (CIntX) to calculate the intrinsic exchange rate constants ( $k_{rc}$ ) of labile protons from backbone, side-chains and termini positions of proteins at defined experimental conditions. CIntX is a fully automated and user-friendly tool in its functions and requires four inputs to achieve the task: PDB file of protein, temperature in Kelvin, pH, and ionic strength. The program accounts the effect of Xaa-Pro peptide bonds, disulfide bonds and ionisable side chains on the  $k_{rc}$  estimations using the 3D structural co-ordinates of the given protein. The program also calculates residue-specific free energies and protection factors of proteins with the only requirement of having NMR-derived  $k_{ex}$  of NHs in the proteins. Estimation of precise  $\Delta G_{HX}$  (free energy of exchange) for proteins at ambient conditions is indispensable to unambiguously address the thermodynamic and kinetic events of the proteins. In these backgrounds, CIntX is a unique tool of this kind for calculating  $k_{rc}$  of labile protons in proteins. The web server of the program is publicly available at <http://sblab.sastru.edu/cintx.html>

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