

# Comparison of Modified 1% Potassium Hydroxide Formol-Ether Concentration Technique with Direct Wet Mount Preparation and Standard Formol-Ether Concentration Technique for Detection of Parasites in Stool

Balakrishna J<sup>1</sup>, Venkateswarlu S<sup>2</sup>, Kusuma Bai<sup>2</sup>, Hussain Saheb S<sup>3</sup>.

<sup>1</sup> Dept. of Microbiology,  
Santhiram Medical College, Nandyal, Andhrapradesh.

<sup>2</sup> Dept. of Microbiology,  
Fathima Institute of Medical Sciences, Kadapa, Andhrapradesh

<sup>3</sup> Dept of Anatomy,  
JJM Medical College, Davangere, Karnataka.

## Abstract

**Objective:** - To compare the diagnostic yield of saline and iodine wet mount preparation with sedimentation techniques (formol-ether concentration technique (FECT) with physiological saline and modified formol-ether concentration technique with 1% potassium hydroxide) of fresh stool sample for diagnosis of human parasitic infections.

**Method:** - Direct wet mount stool slides were examined in saline and iodine preparation and concentration slides were prepared with FECT with physiological saline and FECT with 1% potassium hydroxide (KOH) under low power and high power.

**Results:-** Out of 200 stool samples, 120 (60%) were positive by direct wet mount preparation, 165 (83%) were positive by FECT with physiological saline and 164 (82%) were positive by FECT with 1% KOH. The sensitivity of wet mount and modified 1% potassium hydroxide FECT were 73% and 99.3%, respectively. The positive agreement between FECT and wet mounts (84%) and modified 1% KOH formol-ether concentration techniques (99.7%) were good.

**Conclusion:** - Because of low sensitivity of wet mount preparation; our study conformed that the sedimentation techniques can be utilized for the diagnosis of parasitic infections in humans. The detection rate of FECT with physiological saline and FECT with 1% KOH was not significantly different ( $p > 0.05$ ).

## INTRODUCTION

Human Intestinal parasitic infections persist as a global health problem (1). Laboratory diagnosis of Intestinal parasitic infections depends on identification of intestinal parasites or morphological stages in their life cycle (ova, cyst, trophozoites, or larva) in stool samples (2). Wet mount preparations with saline & iodine is the conventional routine method to detect parasites in clinical samples. Wet mount preparations may fail to detect eggs, cysts or trophozoites when the sample contain parasites in small number. Concentration techniques not only separate fecal debris, it increases the number of parasites in the sediment and making them more visible by removing inorganic and organic debris (3). Formal-ether concentration technique is widely used, for detection of parasites especially trematode eggs and protozoan cyst, with a high content fat in the stool (4). For diagnostic purpose a number of modifications on formal-ether technique have been developed to increase the sensitivity of diagnostic and the effective use of resources (5, 1).

The present study compares with wet mount preparation and a modified 1% potassium hydroxide FECT against the standard procedure FECT method in field conditions (15).

## MATERIALS AND METHODS

Two hundred stool samples were collected from patients who are suffering from persistent diarrhoea with abdominal pain, passage of worms, or allergic reactions cases at Santhiram General Hospital, Nandyal, Andhra Pradesh, India.

Clean, wide mouthed, dry bottles with lids were used to collect all stool samples. All faecal samples were examined by direct wet mount preparation with iodine and saline, and sedimentation techniques using standard procedure FECT comparing to modified 1% potassium hydroxide FECT preparations. Three preparations were used from each sample one for direct wet mount, and second and third for formal-ether technique with normal saline and other with 1% KOH formol-ether concentration technique respectively. Wet mount preparation was carried out as following standard procedure (15). A drop of normal saline and iodine was put at both ends of a clean glass slide, with the help of wooden stick a minute portion of faeces was added in both drops and stirred to make uniform suspension. Coverslips was placed over each suspension separately, with out air bubbles and scanned for parasites for a minimum of 6 minutes before reporting negative. Standard procedure for FECT method was carried out as follows (6, 15). One to two grams of faecal

sample was used for each test. Faeces were mixed in 10ml of water and it was strained through two layers of gauze in a funnel. The filtered solution was centrifuged at 2000 rpm for 2min. Supernatant was discarded. The sediment was suspended in 10ml of physiological saline and again centrifuged, supernatant was discarded. 7ml of 10% formal-saline was added to the sediment allowed for stand for 30 min. Next 3ml of ether was added then tube was shaken vigorously and centrifuged at 2000 rpm for 2min. The supernatant three layers were poured off, the sediment was used for slide preparation. Modified Formal- ether concentration technique with 1% KOH was processed and prepared for microscopy with exactly the same procedure to FECT (15). But 1%KOH was used instead of physiological saline. Both methods were performed by experienced microbiologist. The results of FECT were correlated with the results of wet mount preparation and modified 1% KOH formol-ether concentration technique for all patients.

### RESULTS

It was observed that FECT and modified 1% KOH formol-ether concentration technique proved to be very effective compare to the direct wet mount preparations for detection of parasites table 1. Of 200 stool samples, 120(60%) positive

samples were derived form the wet mount preparation. 165(83%) were derived from the standard formol-ether concentration technique and 164 (82%) were derived from the modified 1% KOH formol-ether concentration technique preparation.

All specimens positive for parasites with the modified 1%KOH formol-ether concentration technique procedure were positive by standard FECT method. 1 of the 165 samples positive by the FECT method were negative by 1% potassium hydroxide FECT method , 45 of the 165 sample positive by the saline FECT method were negative by the wet mount preparation ,44 of the 164 sample positive by the 1% FECT method were negative by the wet mount preparation. Modified 1% potassium hydroxide FECT and wet mount preparations are cross compared with the saline FECT in table 2. The sensitivities of wet mount preparation and modified 1% potassium hydroxide FECT were 73% and 99.3% respectively, and specifications were 100% for each. The positive agreement between the FECT and wet mount preparations was 84% and between standard FECT and modified 1% potassium hydroxide FECT methods was 99.7%, indicating good agreement.

**Table 1**

Cross comparison of the modified 1% KOH formol-ether concentration technique and wet mount methods with standard FECT method.

<i>Parasites</i>	<i>Identification by Wet mount</i>	<i>Identification by standard FECT</i>	<i>Identification by modified1% KOH</i>
<i>Entamoeba histolitica</i>	39	55	55
<i>Etamoeba coli</i>	10	25	25
<i>Giardia intestinalis</i>	20	29	28
<i>Ascaris</i>	10	12	12
<i>Hook worm</i>	18	20	20
<i>Hymenolepis nana</i>	22	23	23
<i>S.stercoralis</i>	1	1	1

**Table 2 –**

Slide results by standard FECT method and Wet mount and modified 1% KOH formol-ether concentration technique methods.

	<i>Standard FECT</i>	<i>Positive</i>	<i>Negative</i>
<i>Wet mount</i>	Positive Negative	120 45	- 35
<i>Modified 1%KOH FECT</i>	Positive Negative	164 1	- 35

### DISCUSSION

Various modified formol-ether concentration techniques have been reported (3,7) for diagnosis of human parasitic infections. The diagnosis of parasites in stool is established by identification of ova and cysts by variety of techniques including direct wet mount preparations, sedimentation method, kato and FECT (8,9,10). Direct wet mount preparation has been widely used to demonstrate parasites in stool (11). However it lacks sensitivity. It could be repeated the examination 4-6 times (11, 12) this may be time consuming for laboratory persons and inconvenient for patients. However, additional use of FECT significantly (13, 14, 12). FECT permits the detections of organisms present even in small numbers: these may be missed by using direct wet mounts. It seems that modified 1% potassium hydroxide FECT can provide a better detection rate (82%) than wet mount preparation (60%). However no statistically significant difference in detection rate between FECT and modified 1% potassium hydroxide FECT could be shown. Standard FECT and modified 1% KOH formol-ether concentration techniques showed greater sensitivity than wet mount preparation. Furthermore, we found that some round worm, eggs, had degenerative surface changes that may have led to inaccurate identification. Though turnaround time for the wet mount preparation was shorter, but parasites may not be detected in mild infections. However both sedimentation technique methods can be used in the preparations of formalin-ether concentration techniques in mild parasitic infections, in which modified 1% potassium hydroxide FECT will reduce stool debris that interfere in test system.

### REFERENCES:

1. Chatterjee KD, Parasitology and helminthology. Sree Saraswaty press Ltd. Calcutta 1978: 207.11.
2. Mims CA, Play fair ill, Toill IMI et al. Medical Microbiology. Fiona Foley Mosby-year Book Ltd., 3rd edition, London, 1933;25:25-30.
3. Jamsai Suwansaksri, Suwannee Nithiuthai, Vivoj Wiwanitkit, Suphan Soogarun and Pennapa Paloitho. The formalin-ether concentration Technique for intestinal parasites: comparing 0.1N Sodium hydroxide with Normal Saline preparations. South East Asian J Trop Med Public Health 2002;33:97-98.
4. Foreyt WJ. Diagnosis parasitology. Vet Clin North Am Small Anim Pract 1989;19:979-1000.
5. Knight WB, Hiatt RA, Cline BL, Ritchie LS. A modification of the formol-ether concentration technique for increased sensitivity in detecting *Schistosoma mansoni* eggs. Am J Trop Med Hyg 1976;25:818-23.
6. Ritchie LS. An ether sedimentation, technique for routine stool examinations Ball us Army Med Dept 1948;8:326.
7. Parija S.C., Bhattacharya, s., Padhan P.& Shiva prakash, M.R. Evaluation of formalin – acetone sedimentation in the concentration on of stool for intestinal parasites. Tropical Doctor 2003; 33:163-164.
8. Chatterjee KD. Parasitology, Protozoology and Helminthology in relation to clinical Medicine. Twelfth edition Calcutta, Sree Saraswaty Press 1980;142-4.
9. Ellen JoB, Peterson LR, FineGold SM. Diagnostic microbiology Ninth edition page 1988;786-7.
10. Abdel Wahab MF. *Schistosoma mansoni* in Egypt. Clin Trop Med Corns Dis 1987;2:371-95.
11. Rabello AI, Rocha RS, de oliveira JP, Katz N, Lambertucci JR. Stool examination and rectal biopsy in the diagnosis and evaluation of therapy of *Schistosoma mansoni*. 1992;34:601-8.
12. Azab M, Al Zayat EA. Evaluation of purified antigens in hemagglutination test for determination of cross reactivities in diagnosis of foscioliasis and schistosomiasis. J Eght Soc Parasitol 1996;26:677-85.
13. Truant AL, Elliott SH, Kelly MI, Smith JH. Comparison of formalinethyl ether Sedimentation, formalin ethyl acetate sedimentation and zinc sulfate flotation technique for detection of intestinal parasite. J Clin Mic 1981;13:882-4.
14. Elliott EE. Schistosomiasis pathophysiology, diagnosis and treatment. Gastroenterol Clin Nrt Am 1996;25:599-625.
15. Cheesbrough M. District laboratory practice in tropical countries, 3<sup>rd</sup> ed. Part one, Cambridge University Press, United Kingdom, 2000;214-215.